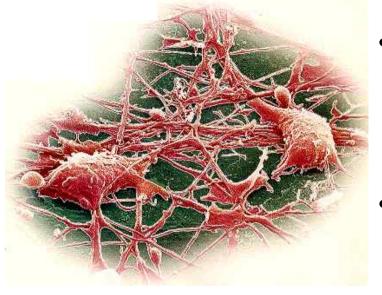
Lesson 12. Content

1. Neural networks.

2. Prediction of secondary structure.

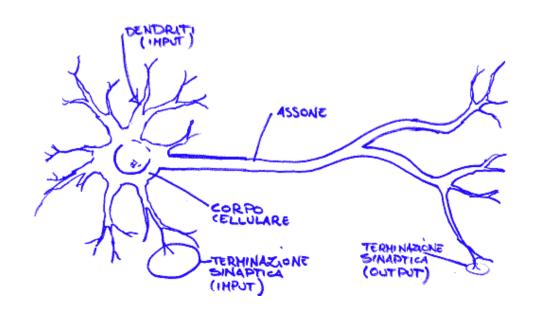
3. Protein contact prediction.

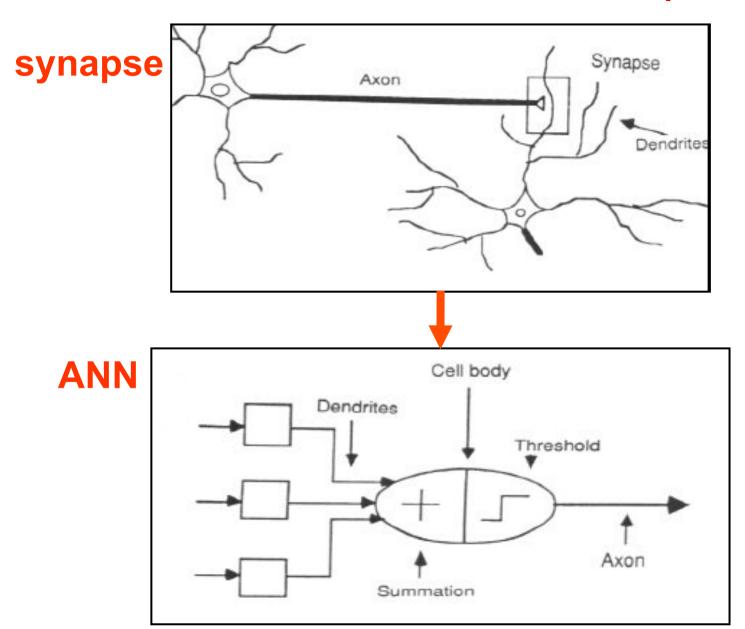
4. 3D structure prediction with Deep Learning.



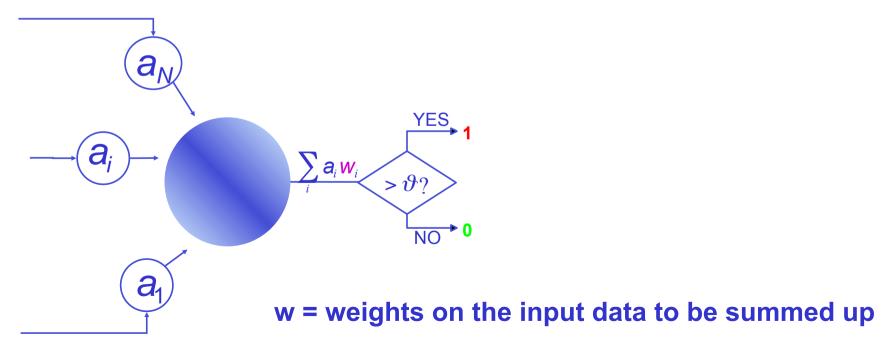
- Computational structures based on (inspired to) the anatomy and physiology of biological neural networks
- Initially developed to simulate information processing and learning in brain

Physiologically, a neuron receives excitatory and inhibitory stimuli (input) and emits a response signal (output) in case the intensity of the stimulus overcomes a given threshold





- They are an example of machine-learning techniques, whose aim is automatically fitting a model value to a known value as closely as possible
- The algorithm will learn from a set of known examples by iterative changes to its parameters – weights of the input data – until the prediction best fits the reality



- ANNs operate by processing information through "layers"; each layer can have many nodes or units
- The simplest NN is a two-layered network, an input layer and an output layer, called perceptron
- The firing of a node in a NN is simulated by assigning the binary values of 1 or 0 to its output; 1 is assigned when the weighted sum of inputs exceeds a predetermined threshold value

	a ₁	a_2	outputexpected
Ex. 1	1	0.3	1
Ex. 2	1	1	1
Ex. 3	0	8.0	0
Ex. 4	0.5	0.4	0

$$\sum_{i} a_{i} w_{i} > \theta \implies YES (1) \quad \sum_{i} a_{i} w_{i} < \theta \implies NO (0)$$

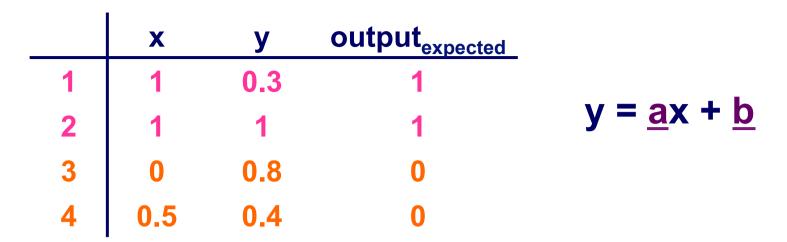
One of the solutions: $w_1 = 1$, $w_2 = 0.5$, $\theta = 0.9$

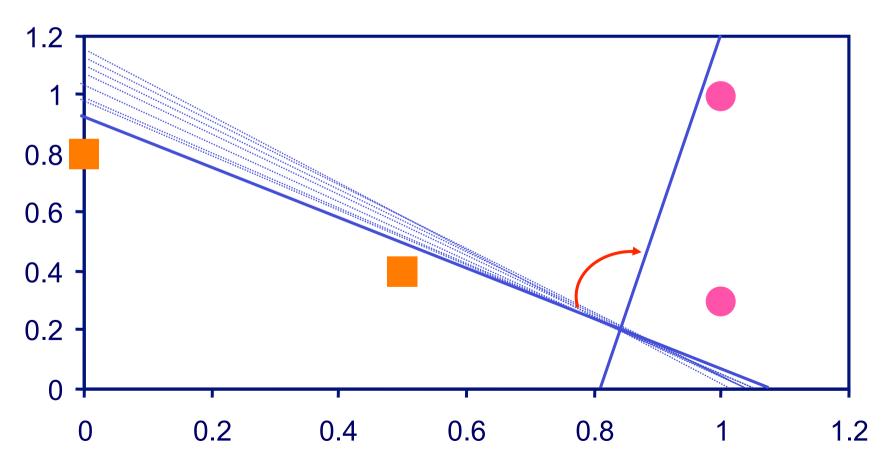
Ex. 1:
$$a_1^*w_1 + a_2^*w_2 = 1*1 + 0.3*0.5 = 1.15 (>0.9) \rightarrow 1$$

Ex. 2:
$$a_1^*w_1 + a_2^*w_2 = 1*1 + 1*0.5 = 1.5 (>0.9) \rightarrow 1$$

Ex. 3:
$$a_1^* w_1 + a_2^* w_2 = 0^*1 + 0.8^*0.5 = 0.4 (<0.9) \rightarrow 0$$

Ex. 4:
$$a_1^*w_1 + a_2^*w_2 = 0.5*1 + 0.4*0.5 = 0.7 (<0.9) \rightarrow 0$$

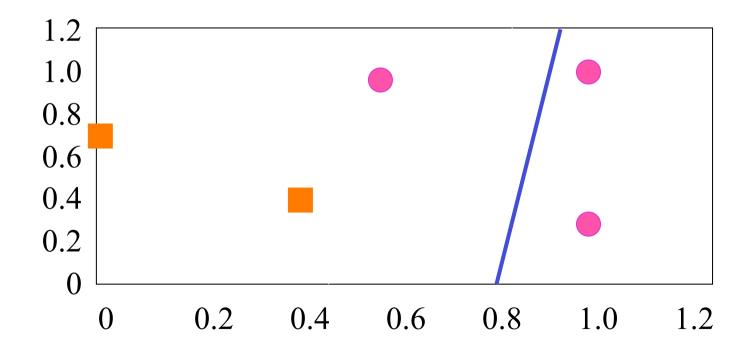




Example of a 2D network

We assign 2 values (coordinates, a_i) to each point & associate a positive or negative output

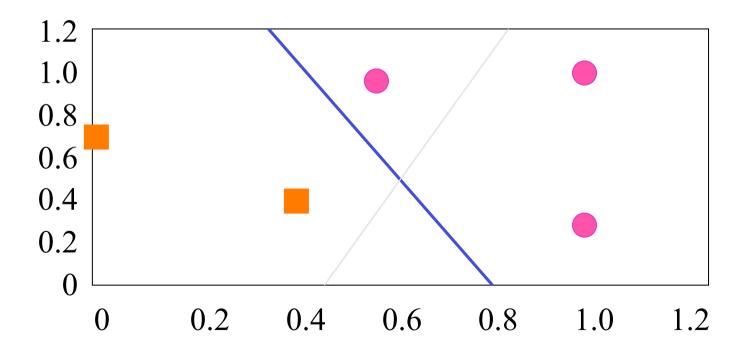
$$X=\sum a_i W_i$$



Example of a 2D network

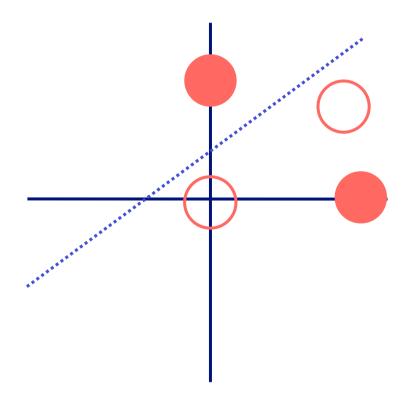
1) We assign 2 values (coordinates, a_i) to each point & associate a positive or negative output

$$X = \sum a_i W_i$$



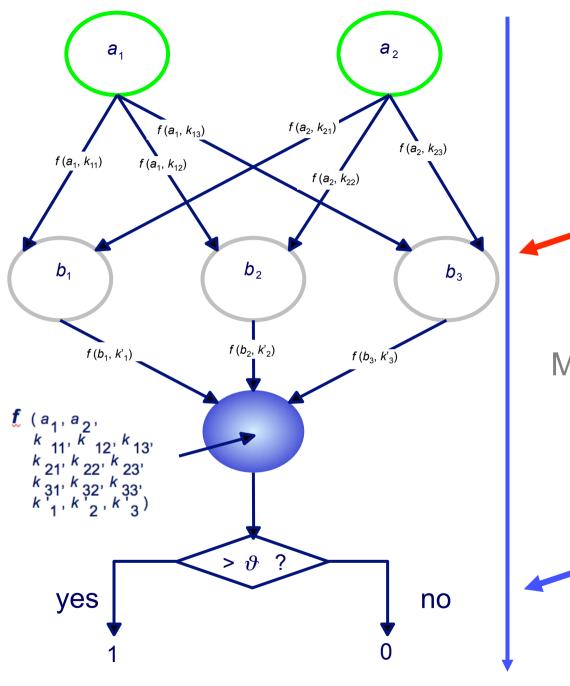
a neural networks can learn from its own mistakes

What function best discriminates between ● & ○ ?



A simple ANN would find at most a dashed straight line

→ We need a more complex network, by introducing an hidden layer



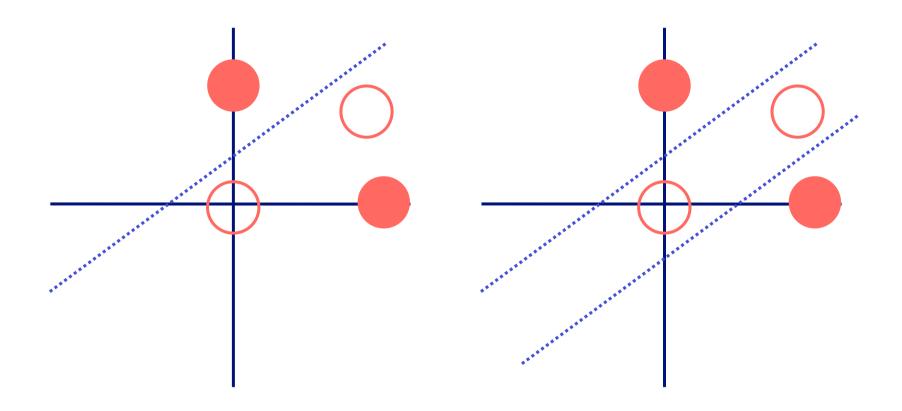
Hidden layer



More parameters to be optimized

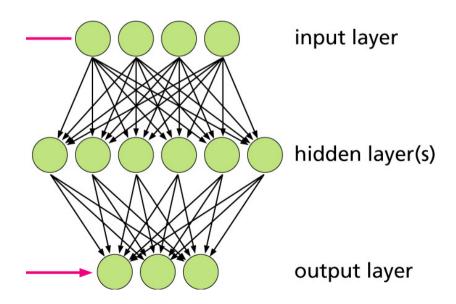
Feed-forward ANN (direction)

Solution



A straight line is not enough to solve the problem, we need two!

- A more complex and more common NN is one that has one or more layers between the input and output ones, the so-called hidden layers
- The hidden layers perform nonlinear transformations of the inputs entered into the network, because there is more than one path to the output node



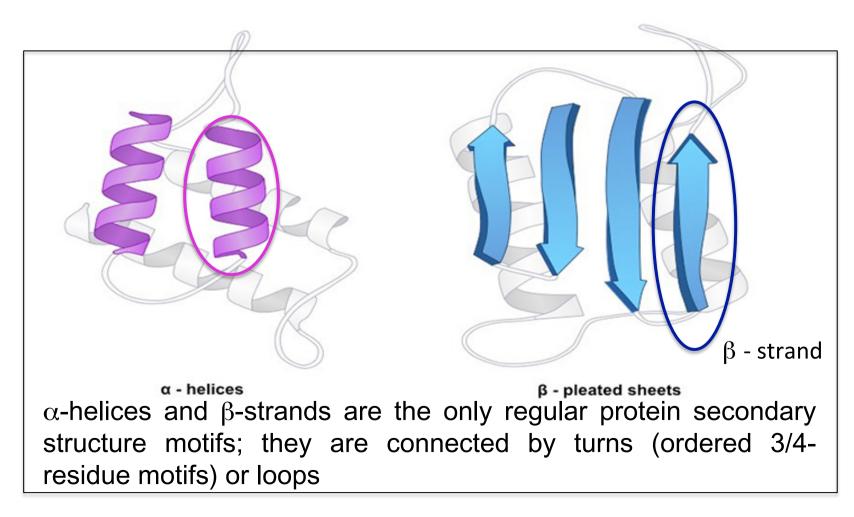
What function best discriminates between • (positives) &

(negatives)? **Calculation of:** TP, FP, TN, FN

No matter how sophisticated the network is, it will always generate some incorrect predictions (FP & FN)

All statistical methods need a validation to be confidently used

 NNs have been widely used in Bioinformatics for the prediction of the secondary structure (SS) of proteins



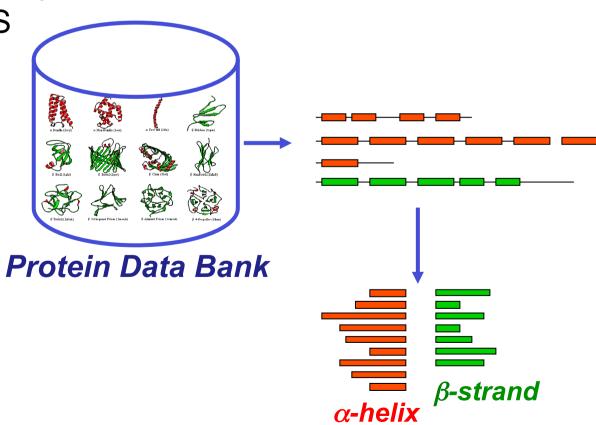
- Application of NNs to the prediction of the protein secondary structure is ideal for at least two reasons:
 - The NN prediction is context-dependent, i.e. different positions in the sequence (or alignment) can have a different relevance (weight) for the prediction



 Application of NNs to the prediction of the protein secondary structure is ideal for at least two reasons:

2. Many examples to learn from are available for the

protein SS



Defining the protein SS: DSSP (Dictionary of protein secondary structure)

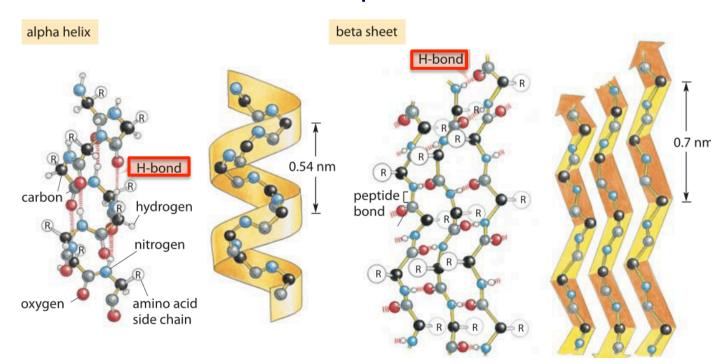
http://swift.cmbi.umcn.nl/gv/dssp/

DSSP-software: assigns the SS according to hydrogen-

bond patterns

DSSP-database: contains SS assignments (plus more info)

for all the protein entries in the PDB.



Defining the protein SS: DSSP (Dictionary of protein secondary structure)

The DSSP code

- H = alpha helix
- **B** = residue in isolated beta-bridge
- E = extended **strand**, participates in **beta-sheet**
- G = 3-helix (3/10 helix)
- I = 5 helix (pi helix)
- T = hydrogen bonded turn
- **S** = bend
- Blank = loop or irregular

Sequence: MNIFEMLRIDEGLRLKIYKDTEGYYTIGIGHLLT-SLDAAKSELDKAIGRNTNGV

DSSP: HHHHHHHHH EEEEEE TTS EEEETTEE - HHHHHHHHHHHTS TTB

Sequence: ITKDEAEKLFNQDVDAAVRGILRNAKLKPVYDSLDAVRRAALINMVFQMGETGVA

Sequence: GFTNSLRMLQQKRWDEAAVNLAKSRWYNQTPNRAKRVITTFRTGTWDAYK

PDB ID: 103L (hydrolase)

- The input signal for an amino acid is usually a group of 20 units in the input layer; the signals of the input will be all 0 except that representing the particular residue, which will be 1
- Usually the sequence is sampled by a sliding window, with the central residue being that for which the SS is predicted (the input is thus a long string of **0**/**1**: for a 13-res window 13x20 units)

	A	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	Y	W
V	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
E	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
E	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
F	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
L	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
G	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
H	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
C	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
L	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
R	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
I	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
M	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
S	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
C	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

- When using multiple aligned sequences, the input layer signals will be related to sequence profiles based on these alignments
- Information contained in multiple alignments increases the accuracy of prediction, because proteins preserve their SS during evolution

	A	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	Y	W
AAAAV	. 8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	. 2	0	0
D D D E E	0	0	. 6	. 4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
EEEEE	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
FFWFF	0	0	0	0	. 8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	. 2
ILLLL	0	0	0	0	0	0	0	. 2	0	. 8	0	0	0	0	0	0	0	0	0	0
GGGGG	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
н н н н н	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
CCCCC	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
ILLLL	0	0	0	0	0	0	0	. 2	0	. 8	0	0	0	0	0	0	0	0	0	0
K K R K R	0	0	0	0	0	0	0	0	. 6	0	0	0	0	0	. 4	0	0	0	0	0
LIIII	0	0	0	0	0	0	0	. 8	0	.2	0	0	0	0	0	0	0	0	0	0
M M M M M	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
N N S S S	0	0	0	0	0	0	0	0	0	0	0	. 4	0	0	. 6	0	0	0	0	0
CCCCC	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

- The output layer usually consists of 3 units, corresponding to the three alternative conformations to predict (a-helix, b-strand, loop/coil)
- An output like (1, 0, 0) would correspond to a perfect helix prediction; however prediction is usually done based on the highest number in output (see below)

N-terminal... T H I S I S A H I D D E N M E S S A G E ... C-terminal

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hidden layer

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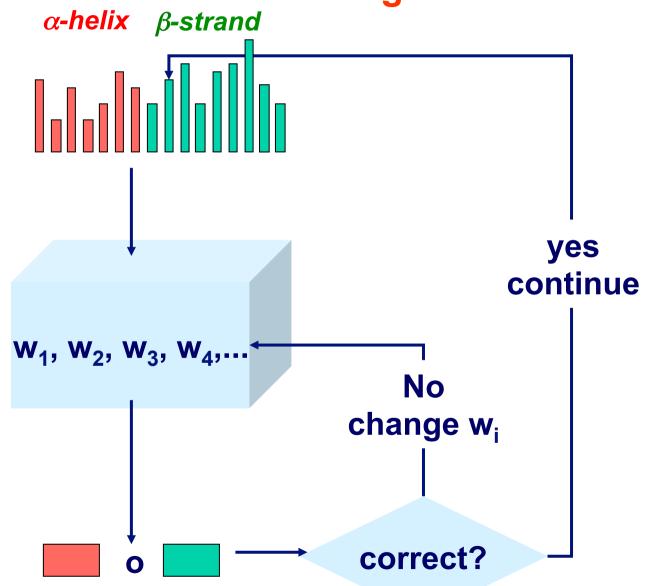
A simplified representation of a multilayer NN:
prediction is made on the central residue of the window (an IIe);
layer nodes receiving signals above a certain value, e.g. the red one, will fire to the output layer (prediction: helix);
confidence of prediction can be related to how close to 1 is the highest number

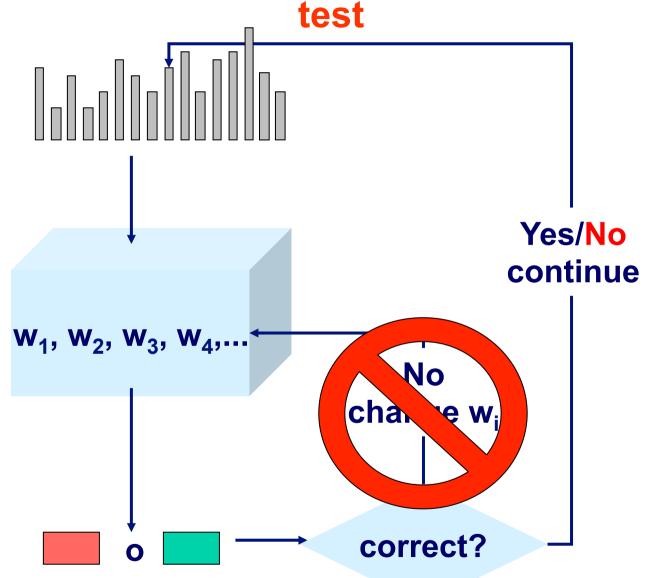
Implementing a NN requires three phases:

 Training: method development using non-homologous protein sequences of known structure

 Test: check of the method on protein sequneces of known structure

Validation: statistical analysis of obtained results

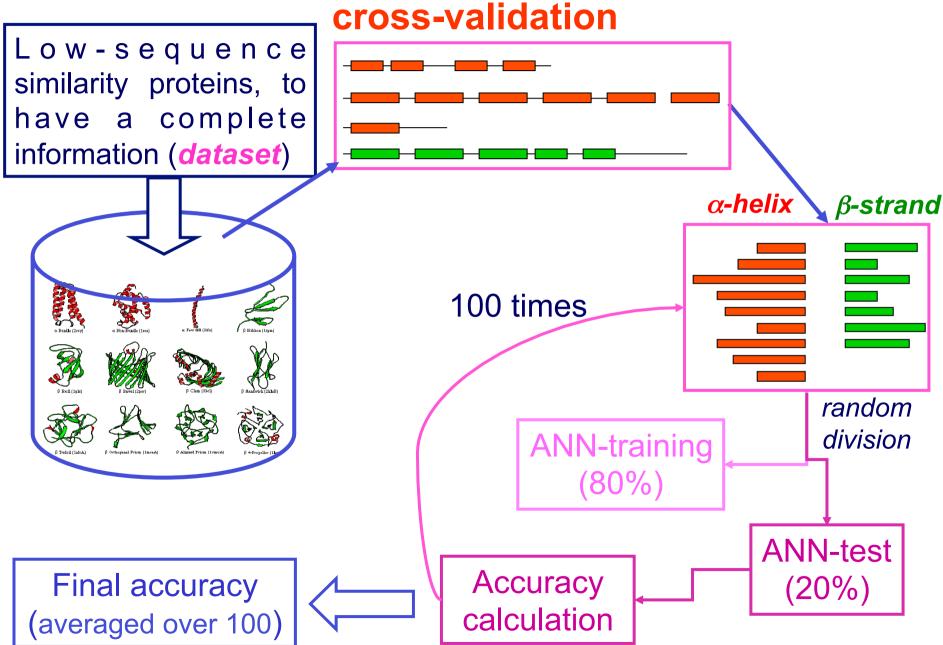




What sequences do we use for the test phase? They must also be structure-known

Validation

- To be reliable, knowledge-based methods <u>must</u> be tested with a rigorous statistics
- The most commonly used validation statistics is the crossvalidation (or jack-knife test)
- From cross-validation results measures of the prediction performance (such as sensitivity, specificity, correlation coefficient etc.) can be calculated, which are universal, therefore comparable and reproducible



Accuracy evaluation parameters

• Q3 = percentage of sequence expected to have a correct SS prediction based on 3-state classification, H-E-C

Q3 =
$$\left(\frac{\text{TP(H)}}{\text{Tot(H)}} + \frac{\text{TP(E)}}{\text{Tot(E)}} + \frac{\text{TP(C)}}{\text{Tot(C)}}\right)^* 100$$

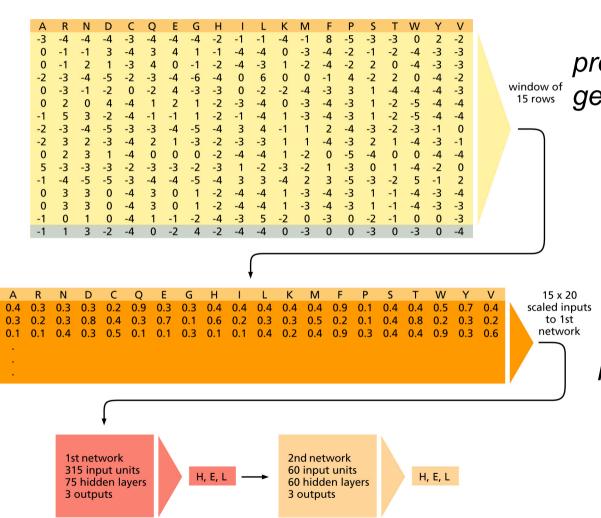
 Matthew's = geometrical mean of the correlation coefficients relative to the three states H-E-C (preferable to Q3)

$$CC_{H} = \frac{TP(H)*TN(H) - FP(H)*FN(H)}{\sqrt{(TP(H)+FP(H))*(TP(H)+FN(H))*(TN(H)+FP(H))*(TN(H)+FN(H))}}$$

$$CC_M = \sqrt[3]{CC_H * CC_E * CC_C}$$
 (geometrical mean)

Server online: PSIPRED

Several SS prediction servers based on NNs are available, including **PSIPRED** and **PHDsec**



PSIPRFD dual network prediction: first a raw profile generated by PSI-BLAST is taken and scaled to a 0-1 range. A window of 15 elements is fed to the 1st network, which performs the initial SS prediction using various residue parameters. This initial prediction is fed into a 2nd NN where it is filtered to produce the final threestate SS prediction

An example...

>gi|15595724|ref|AAG03916.1| transcriptional regulator Dnr [Pseudomonas aeruginosa PA01]

MEFQRVHQQLLQSHHLFEPLSPVQLQELLASSDLV

NLDKGAYVFRQGEPAHAFYYLISGCVKIYRLTPEG

QEKILEVTNERNTFAEAMMFMDTPNYVATAQAVVP

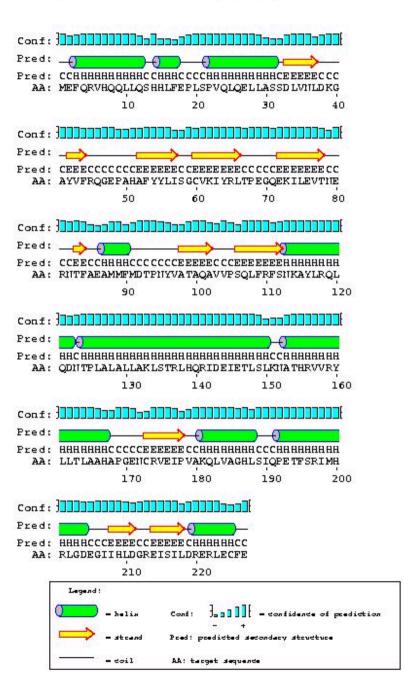
SQLFRFSNKAYLRQLQDNTPLALALLAKLSTRLHQ

RIDEIETLSLKNATHRVVRYLLTLAAHAPGENCRV

EIPVAKQLVAGHLSIQPETFSRIMHRLGDEGIIHL

DGREISILDRERLECFE

PSIPRED results



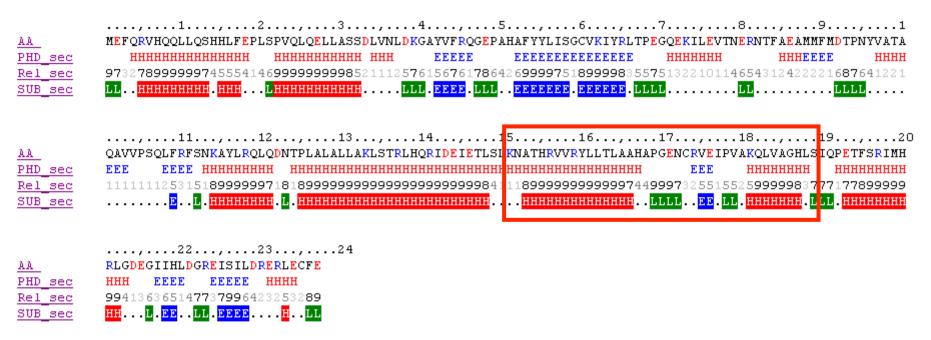
An example...

>gi|15595724|ref|AAG03916.1| transcriptional regulator Dnr [Pseudomonas aeruginosa PA01]

MEFQRVHQQLLQSHHLFEPLSPVQLQELLASSDLVNLDKGAYVFRQGEPAHAFYYLISGCVKIYRLTPEG
QEKILEVTNERNTFAEAMMFMDTPNYVATAQAVVPSQLFRFSNKAYLRQLQDNTPLALALLAKLSTRLHQ
RIDEIETLSLKNATHRVVRYLLTLAAHAPGENCRVEIPVAKQLVAGHLSIQPETFSRIMHRLGDEGIIHL
DGREISILDRERLECFE

PHDsec results

PHD results (normal)



Confidence scores

To each predicted sequence position a confidence score is associated which indicates the probability of the prediction to be correct

Dnr da Pseudomonas aeuruginosa

...LLTLAAHAPGENCRVEIPVAKQ...



...LLTLAAHAPGENCRVEIPVAKQ...

...HHHHHHhcCCCceEEEEeCCHH...

...9998874499802899725989...

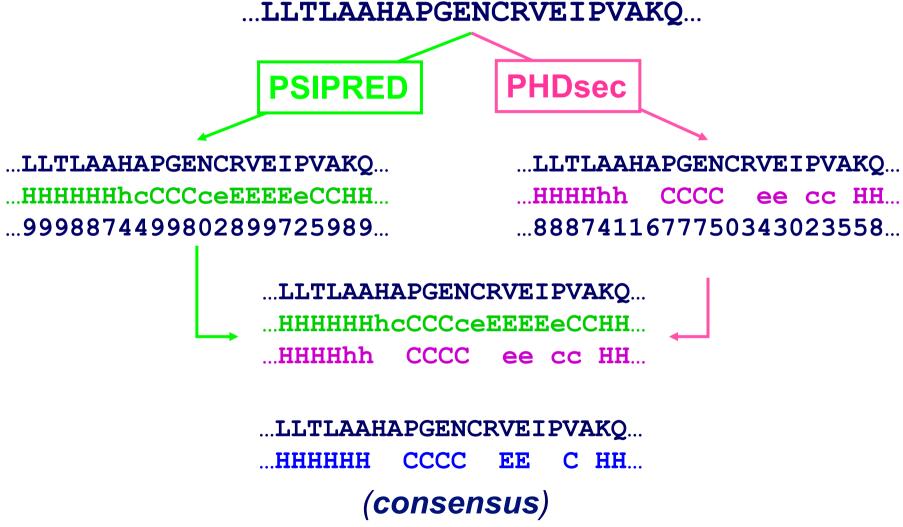
...LLTLAAHAPGENCRVEIPVAKQ...

...HHHHhh CCCC ee cc HH...

...8887411677750343023558...

Metaserver: resources exploiting and combining the best SS prediction methods and improve their performance

Dnr da Pseudomonas aeuruginosa

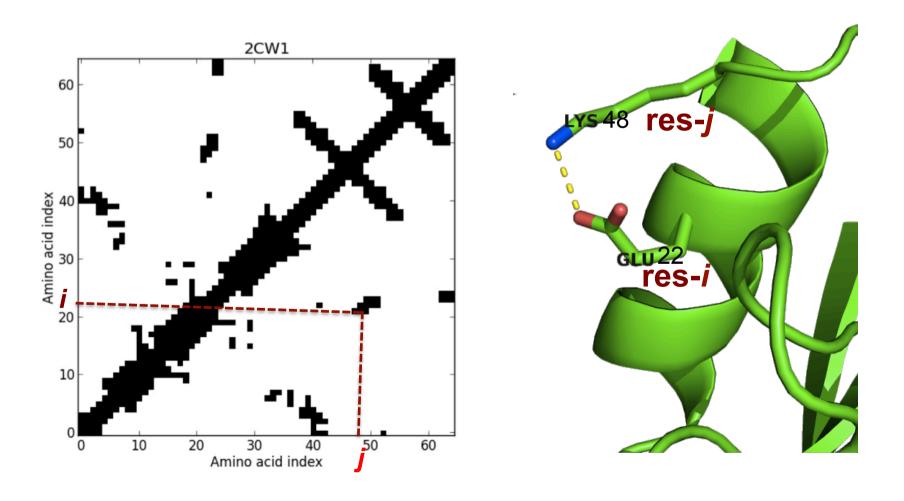


Accuracy of NN-based methods for the prediction of protein secondary structure can be vary high, up to 90-95%

Accuracy for a given query depends on the availability of homologs for it, i.e. on the availability of evolutionary information...

Protein contact map

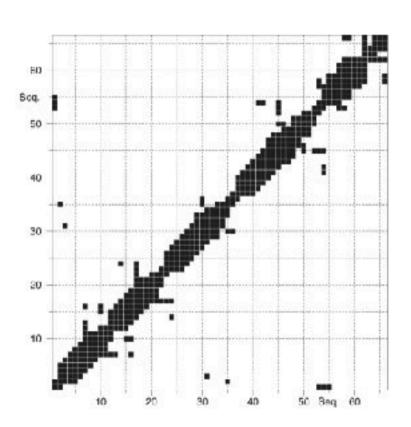
A protein contact map is a 2D representation of a protein where a black dot is present at the cross-over of two residues (i and j), if they are closer than a given cut-off distance (usually 6 Å).



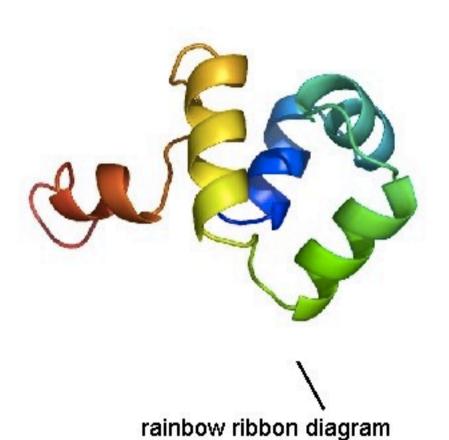
Protein contact map

In this example only contacts between residues with their $C\alpha$ within 6 Å are considered

map of $C\alpha$ - $C\alpha$ distances < 6 Å



Both axes are the sequence of the protein



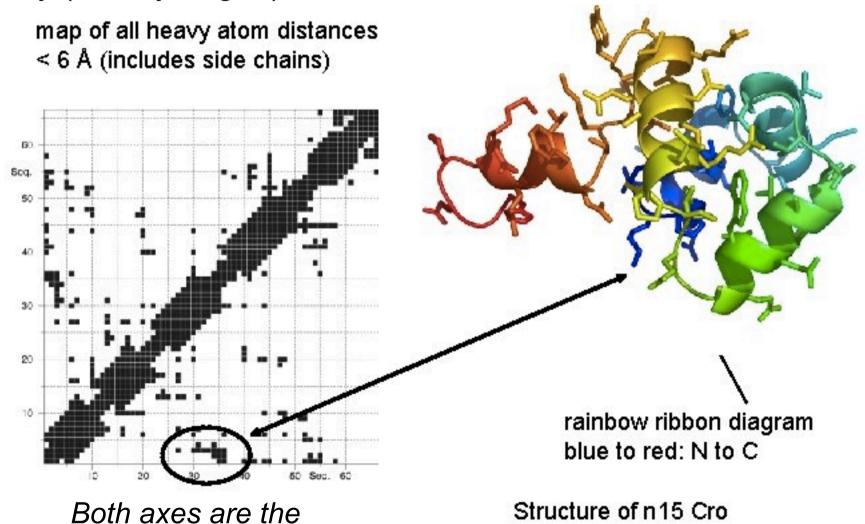
Structure of n15 Cro

blue to red: N to C

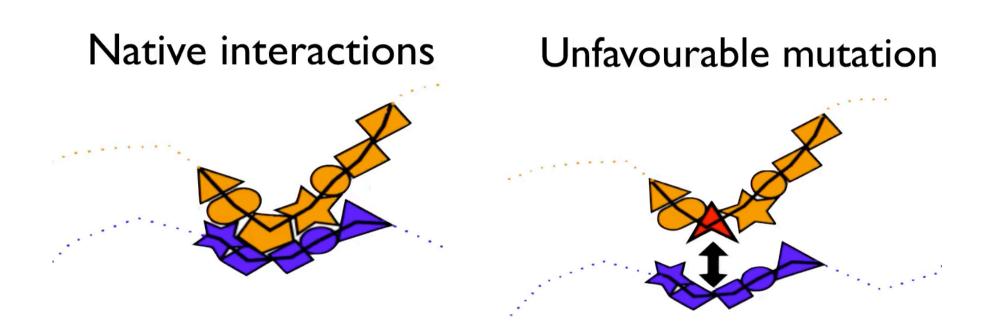
Protein contact map

sequence of the protein

In this example contacts between residues with any of their heavy (non-hydrogen) atoms within 6 Å are considered

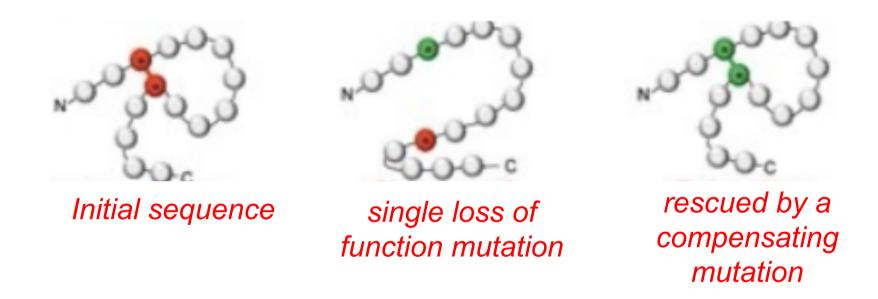


Residue-residue contacts



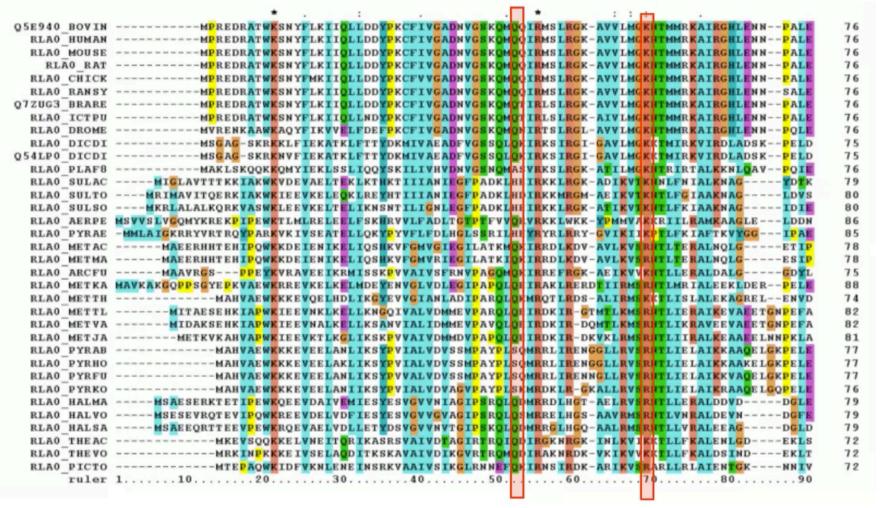
Residue which are in close contact tend to be complementary in shape and properties

Residue-residue contacts

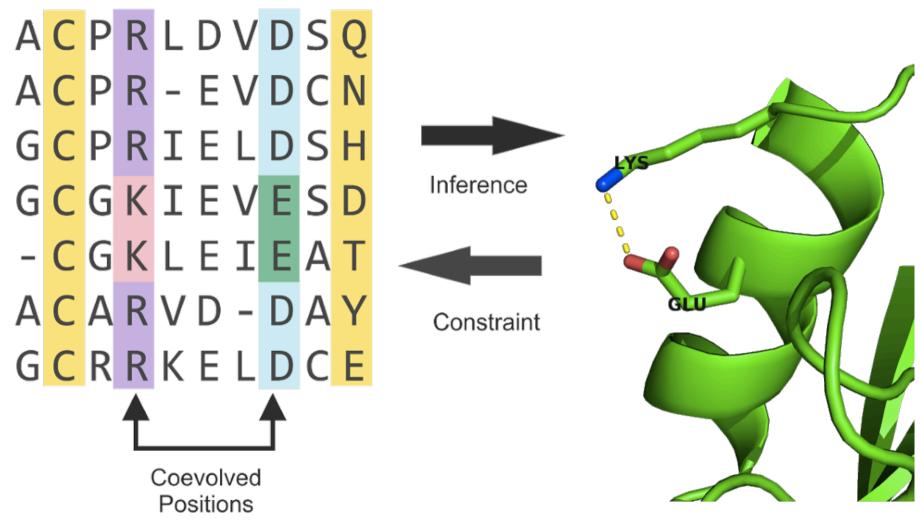


Residue which are in close contact tend to be complementary in shape and properties

If one of them gets mutated, a compensating mutation will most probably occur to the other amino acid involved in the contact



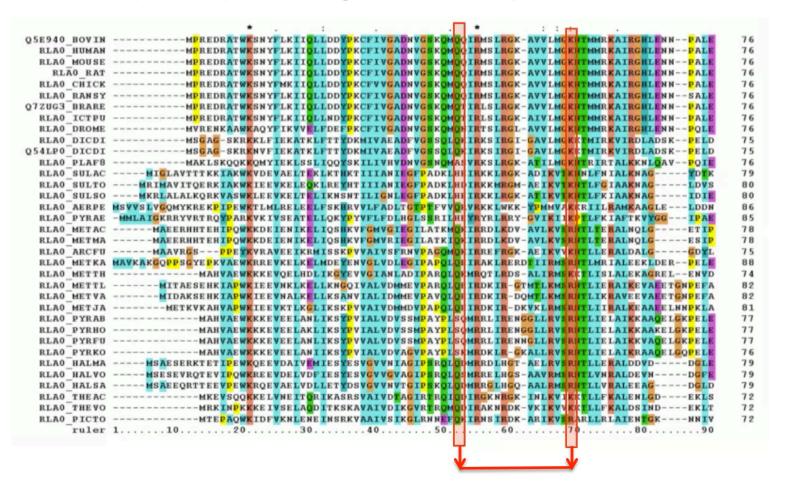
The theoretical basis for residue-residue contact prediction is that residues which are in contact tend to co-evolve, in order to stay nicely complementary



The theoretical basis for residue-residue contact prediction is that <u>residues which are in contact tend to co-evolve</u>, in order to stay nicely complementary

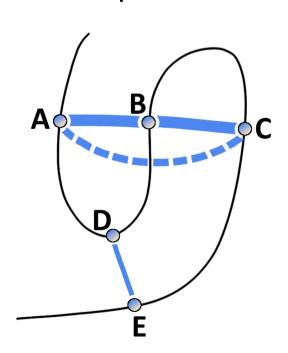
The MSA of a protein family comprises homolog sequences from a common ancestor aligned relative to each other

Therefore, **compensatory mutations** in MSA columns can be **used to infer spatial proximity** of residue pairs



Early contact prediction methods used local pairwise statistics to infer contacts considering pairs of amino acids as statistically independent from others

The traditional covariance approaches suffered from high false positive rates because of their inability to cope with <u>transitive</u> <u>effects</u> that arise from chains of correlations between multiple residue pairs



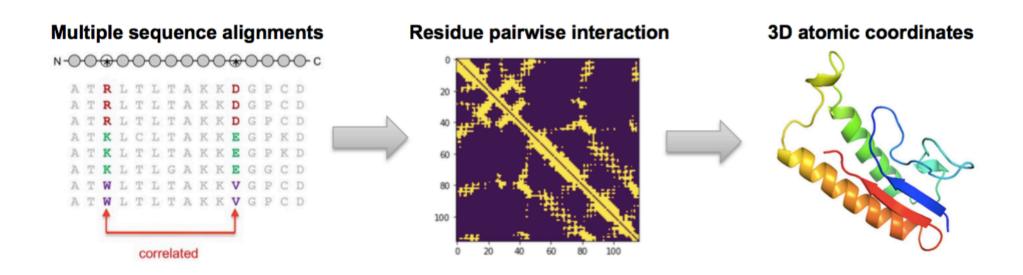
Considering three residues A, B and C, where A physically interacts with B and B with C, strong statistical dependencies between pairs (A,B) and (B,C) can induce strong indirect signals for residues A and C, although they are not physically interacting, which can be even larger than signals of other directly interacting pairs (D,E) and thus lead to false predictions

To deal with this, first a global statistical model that made predictions for a single residue pair while considering all other pairs in the protein was developed, which represented a huge leap forward

Then, machine-learning based methods, including neural networks, have emerged that extract features from MSAs in order to learn associations between input features and residue-residue contacts

Sequence features used in input typically include predicted solvent accessibility, predicted secondary structure, contact potentials, conservation scores, pairwise coevolution statistics, etc.

When residue pairwise interactions (contact maps) are predicted based on coevolution, i.e. on the MSA obtainable for a protein, they can be used for predicting its 3D structure



Deep learning

Deep learning methods are machine learning (ML) methods based on artificial neural networks (ANNs), also named deep neural networks (DNNs)

The adjective "deep" in deep learning refers to the use of multiple layers in the network

Since the 2010s, advances in ML algorithms and computer hardware have led to more efficient methods for training DNNs that contain many layers of non-linear hidden units

Artificial Intelligence: Mimicking the intelligence or behavioural pattern of humans or any other living entity. Machine Learning: A technique by which a computer can "learn" from data, without using a complex set of different rules. This approach is mainly based on training a model from datasets. Deep Learning: A technique to perform machine learning inspired by our brain's own network of neurons.

Deep learning: common applications

Within science

DNNs have been successfully applied to predict the biomolecular target of a drug, to detect toxic effects of environmental chemicals in nutrients, household products and drugs, etc.

Outside science

Fraud detection

Customer relationship management systems

Computer vision

Vocal Al

Natural language processing

Autonomous vehicles

Supercomputers

Investment modeling

E-commerce

Siri, Alexa, Cortana, Google Assistant, etc., are all very popular applications of Deep Learning

Deep learning: limitations

Deep learning and neural networks in general may have two main limitations:

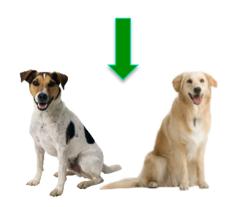
overfitting, i.e. the production of an analysis that corresponds too closely or exactly to a particular set of data, and may therefore fail to fit to additional data or predict future observations on unseen data; an overfitted model contains more parameters than can be justified by the data. It can be a consequence of the training data being incomplete and redundant

computational time, the more sophisticated is the network the more CPU time it will require

Data diversity (heterogenicity) vs overfitting



Training Data (the most diverse the better)



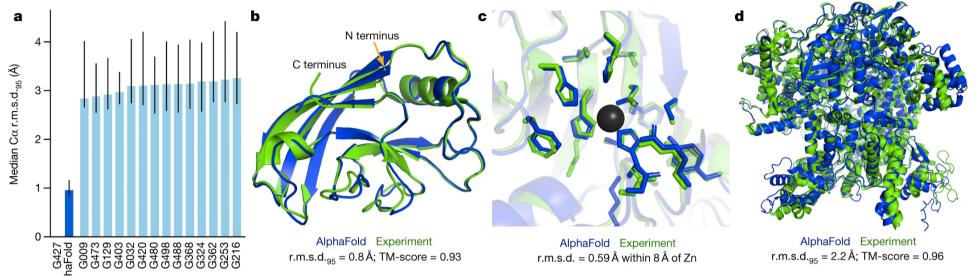


risk of overfitting: data correspond to a specific dog breed

Future observations to be predicted

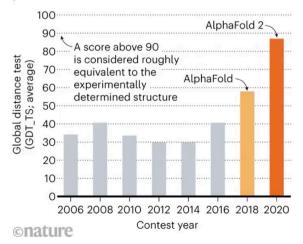
Example: dog recognition

AlphaFold2: the structure prediction miracle Performance on the CASP14 dataset (n = 87 protein domains)

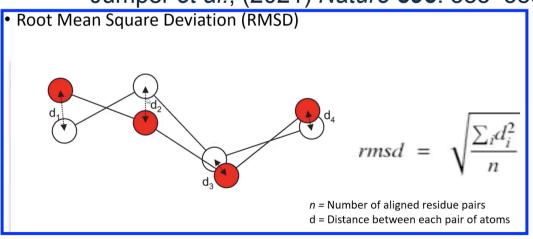


STRUCTURE SOLVER

DeepMind's AlphaFold 2 algorithm significantly outperformed other teams at the CASP14 proteinfolding contest — and its previous version's performance at the last CASP.



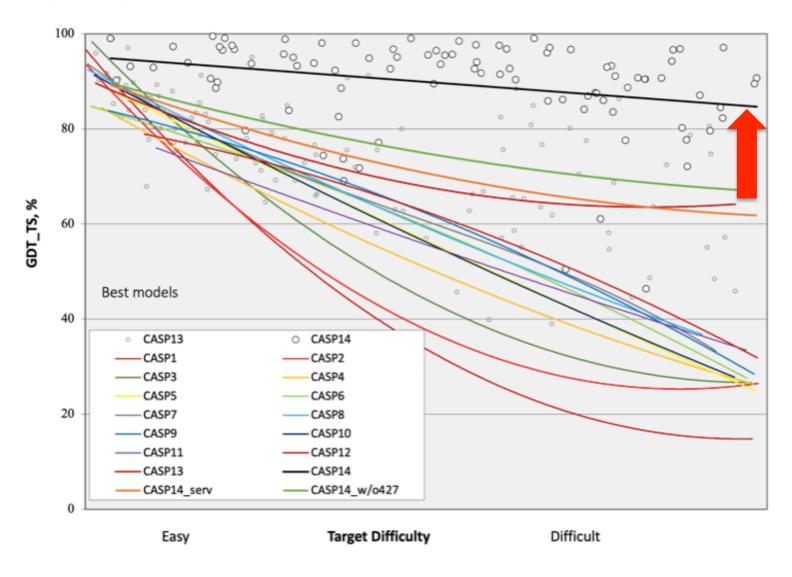
Jumper et al., (2021) Nature **596**: 583–589

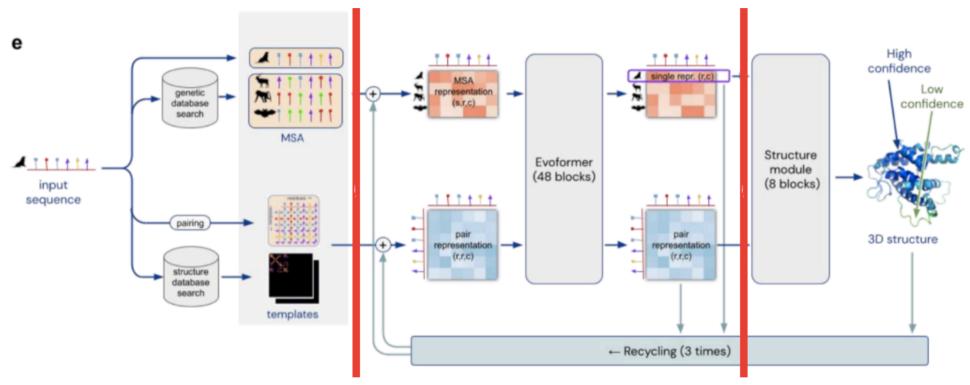


GDT_TS: percentage of corresponding α-carbons within a 4 Å distance

AlphaFold2: the structure prediction miracle Performance on the CASP14 dataset (n = 87 protein domains)

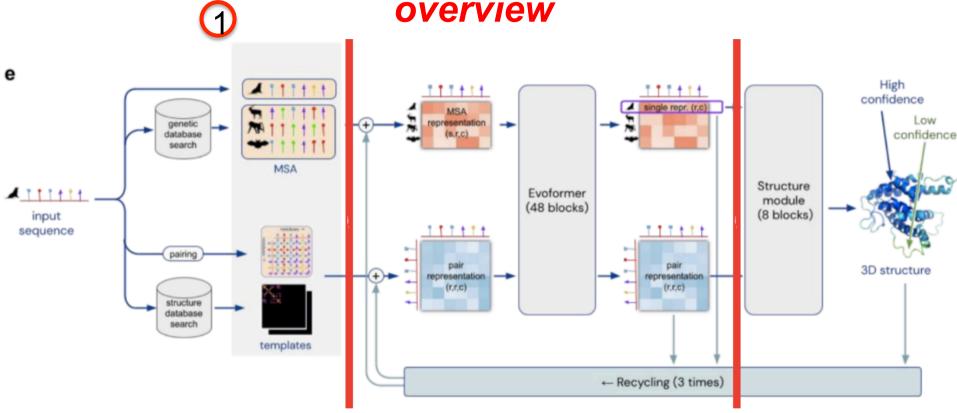
The leap in performance in CASP14





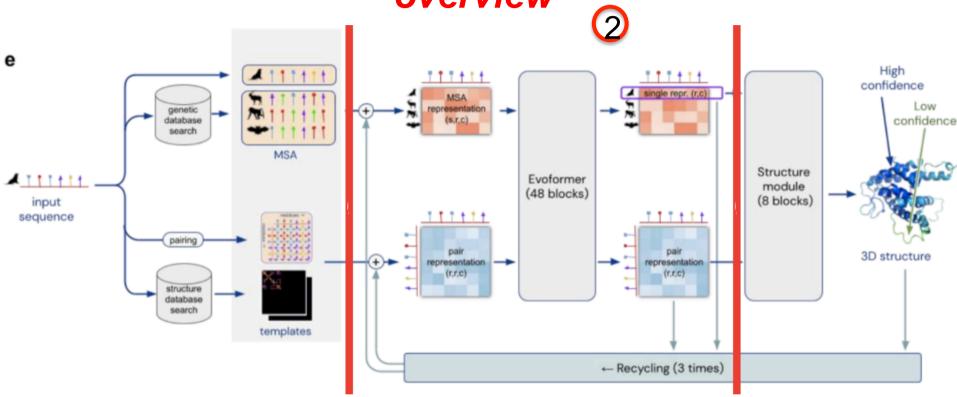
Jumper et al., (2021) Nature **596**: 583–589

https://www.blopig.com/blog/2021/07/alphafold-2-is-here-whats-behind-the-structure-prediction-miracle/



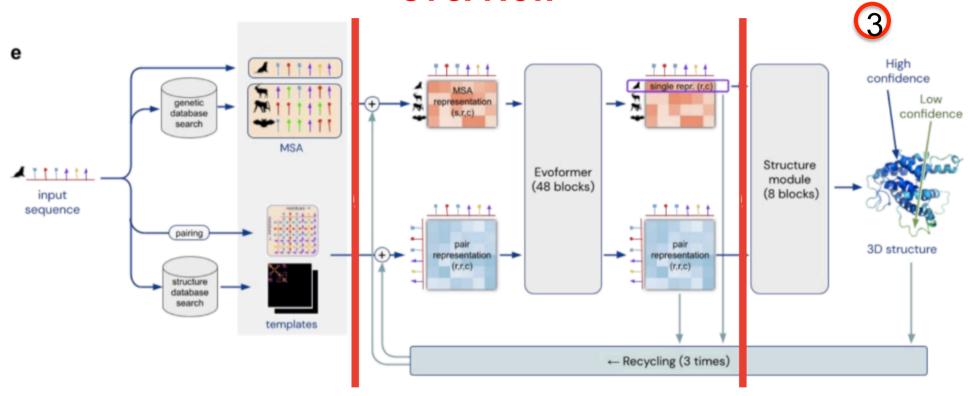
First, AlphaFold 2 uses the input amino acid sequence to query several databases of protein sequences, and constructs a multiple sequence alignment (MSA) highlighting the parts of the sequence that are more likely to mutate and possible correlations

It also tries to identify proteins that may have a similar structure to the input ("templates"), and constructs an initial representation of the structure (the "pair representation"), i.e. a model of which amino acids are likely to be in contact with each other



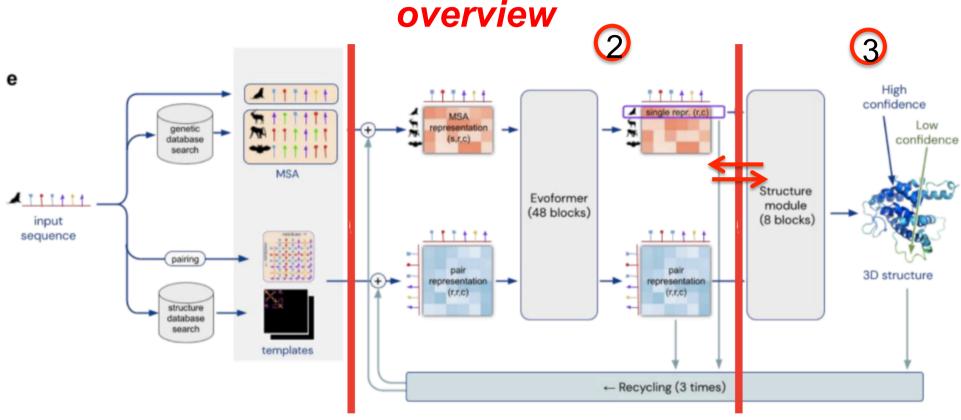
Then, AlphaFold 2 takes the MSA and the templates, and passes them through a transformer (*Evoformer, a neural network*), sort of an "oracle" that can quickly identify which pieces of information are more informative

The objective of this part is to refine the representations of the MSA and the pair interactions, and to iteratively exchange information between them. This process is organised in blocks that are repeated iteratively (48 blocks in the published model)



The last part is the structure module. This piece of the pipeline (again *a neural network*) takes the refined "MSA representation" and "pair representation", and leverages them to construct a three-dimensional model of the structure

This network does not use any optimization algorithm: it generates a final 3D structure, including side chains, in a single step

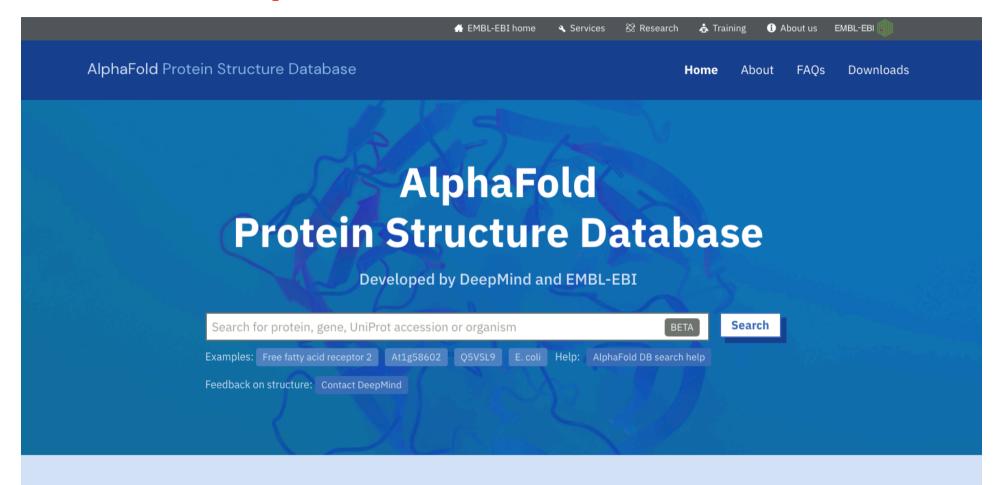


The model works iteratively. After generating a final structure, it will take all the information (i.e. MSA representation, pair representation and predicted structure) and pass it back to the beginning of the Evoformer blocks

This allows the model to refine its predictions

https://static-content.springer.com/esm/art%3A10.1038%2Fs41586-021-03819-2/MediaObjects/41586_2021_3819_MOESM5_ESM.mp4

AlphaFold2: the DataBase

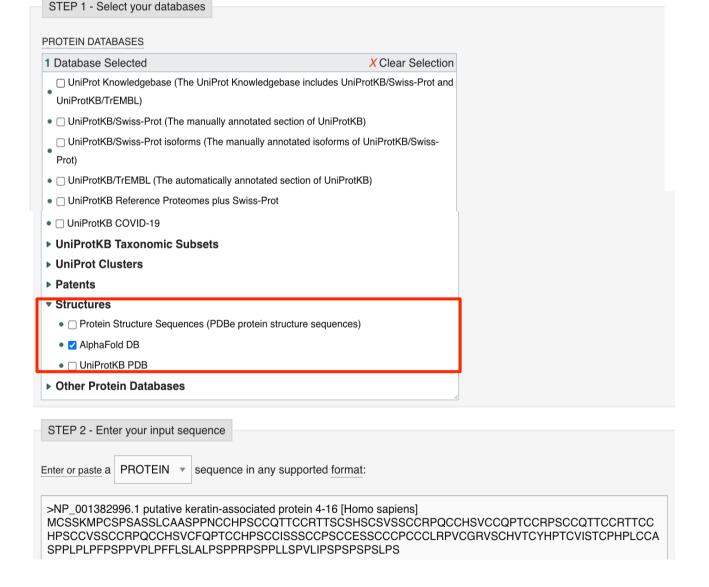


AlphaFold DB provides open access to <u>over 200 million protein</u> <u>structure predictions</u> to accelerate scientific research.

Also available in FASTA

Protein Similarity Search

This tool provides sequence similarity searching against protein databases using the FASTA suite of programs. FASTA provides a heuristic search with a protein query. FASTX and FASTY translate a DNA query. Optimal searches are available with SSEARCH (local), GGSEARCH (global) and GLSEARCH (global query, local database).



Cytochrome c oxidase subunit 1

AlphaFold structure prediction

An example...

Download PDB file mmCIF file Predicted aligned error

Note: We have recently updated the PAE JSON format, please refer to our FAO for a description of the updated format.

NEW Feedback on structure Looks great Could be improved

Cytochrome c oxidase subunit 1

Information

Gene mt-co1

Protein

Source organism Carassius auratus (Goldfish) go to search [5]

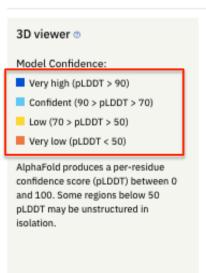
UniProt 078681 go to UniProt ☑

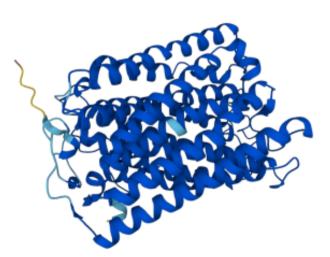
Experimental structures None available in the PDB

Biological function Component of the cytochrome c oxidase, the last enzyme in the mitochondrial electron transport chain which drives oxidative phosphorylation.

The respiratory chain contains 3 multisubunit complexes succinate dehydrogenase (complex II, CII), ubiquinol-cytochrome c oxidoreductase (cytochrome b-c1 complex, complex III, CIII) and cytochrome c oxidase (complex IV, CIV), that cooperate to transfer electrons derived from NADH and succinate to molecular oxygen, creating an electrochemical gradient over the inner membrane that drives ... + [show more] go to

UniProt 12







AlphaFold2 confidence score

The AlphaFold2 confidence score is the **pLDDT**: **predicted Local-Distance Difference Test**

- ◆ Regions with **pLDDT > 90** are expected to be modelled to <u>high accuracy</u>. These should be suitable for any application that benefits from high accuracy (e.g. characterizing binding sites)
- Regions with **pLDDT between 70 and 90** are expected to be modelled well (a generally good backbone prediction)
- Regions with **pLDDT between 50 and 70** are <u>low</u> <u>confidence</u> and should be treated with caution.
- Regions with **pLDDT < 50** should not be considered, they are most probably <u>unstructured</u> (disordered) in physiological conditions or only structured as part of a complex

HCG2042993

Another example...

 \wedge

AlphaFold structure prediction

Download PDB file mmCIF file Predicted aligned error

NEW Feedback on structure Looks great Could be improved

Information

Protein HCG2042993
Gene KRTAP4-16

Source organism Homo sapiens (Human) go to search &

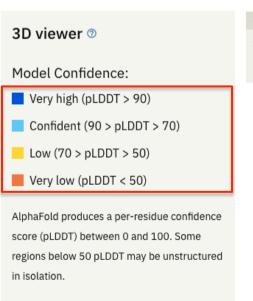
UniProt G5E9R7 go to UniProt

Experimental structures None available in the PDB

Biological function In the hair cortex, hair keratin intermediate filaments are embedded in an interfilamentous matrix, consisting of hair keratin-associated proteins

(KRTAP), which are essential for the formation of a rigid and resistant hair shaft through their extensive disulfide bond cross-linking with abundant

cysteine residues of hair keratins. The matrix proteins include the high-sulfur and high-glycine-tyrosine keratins. go to UniProt &





Lesson 12. Content

- 1. Neural networks (NNs). Mimic physiological NNs. Part of Artificial Intelligence (AI) methods. Can learn from their own errors. Need many diverse examples with a known answer to learn (to be trained) from; when complex (multilayers etc.) need high computational power
- 2. Prediction of secondary structure. Highly efficient. Performed based on NNs since at least two decades.
- 3. Protein contact prediction. Recently recognized as an efficient basis for protein 3D structure prediction. Exploits evolutionary info through co-evolution in MSAs.
- 4. 3D structure prediction with Deep Learning. Come into field in the last few years, set a revolution in it. Reaches experimental-like accuracy in most cases.